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# The Biomechanics and Biology of the Spinal Degenerative Cascade

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More than 25 years-ago, Kirkaldy-Willis et al presented the concept of a cascade of spinal motion segment degeneration invoking progressive wear of the intervertebral disc and facet joints<sup>1</sup>. (Fig.1) The authors emphasized the interdependence of the disc and facet joints for normal spinal function and described how derangement or injury to either of these articulations, leads to abnormal forces and impairment of the other, the so called “tripod” effect. They further described the morphologic features of spinal degeneration and postulated how these might be associated with various clinical syndromes. Although insightful, this algorithm was quite mechanistic and, in keeping with the times, highlighted biomechanical disturbances associated with degeneration of the motion segment. Over the decades since, we have come to appreciate that spinal degeneration involves a complex interplay of biologic and biomechanical events that are predisposed to by genetic factors and modulated by environmental influences.

Degeneration of the spine is an inevitable consequence of aging. Miller et al reported an increase in disc degeneration from 16% at age 20 to approximately 98% at age 70 years based on macroscopic disc degeneration grades of 600 autopsy specimens. Interestingly, the authors noted that lumbar disc degeneration was already present in 11- to 19-year old males and 10 years later in females<sup>2</sup>. Although spinal degeneration is inevitable with aging, it is typically asymptomatic. A more recent MRI study has also achieved similar results<sup>3</sup>.

Kirkaldy-Willis et al postulated that injury or repetitive strain to the facet joint is a cardinal event in the spinal degenerative sequence<sup>1</sup>. More recently, the intervertebral disc

has received considerable attention as the source of initial spinal motion segment dysfunction. Butler et al suggested that disc degeneration likely predates facet arthrosis based on a CT and MRI study<sup>4</sup>. The authors noted that in 68 patients (330 discs / 390 facet joints) there were 144 degenerated discs and 41 levels with facet osteoarthritis. Disc degeneration without facet osteoarthritis was found at 108 levels, while all but one of 41 levels with facet degeneration also had disc degeneration<sup>4</sup>.

The wide spread acceptance that spinal pain often originates from the intervertebral disc is further evidenced by the host of diagnostics (including discography) and therapeutic interventions directed towards the disc. Most treatments for so-called “painful discs” have however met with inconsistent clinical outcomes<sup>5</sup>, probably reflecting a relatively unsophisticated approach to understanding spinal pain. Recent data supporting the idea of facet (zygapophyseal) joint mediated pain have come from studies of patients sustaining cervical whiplash injuries. Lord et al evaluated cervical zygapophyseal joint pain after whiplash in a diagnostic double-blind study using placebo-controlled local anesthetic blocks. 68 patients with a predominant complaint of neck pain and headaches after a whiplash injury were evaluated. The authors noted that among patients with dominant headache, comparative blocks revealed that the prevalence of C2-3 zygapophyseal joint pain was 50%. Overall, the prevalence of cervical zygapophyseal joint pain was 60% (95% confidence interval, 46-73%)<sup>6,7</sup>. These studies further support the complex interplay of the IVD and facet joints in health and disease of the spine.

Our understanding of spinal degeneration has advanced as we have appreciated that the degenerative cascade involves interplay of both biologic and biomechanical factors. Biochemical events are important in the pathogenesis of the degenerative process as well as in the pain-signaling pathways responsible for the clinical features of the condition. As we better appreciate the biologic aspects of spinal degeneration, less-invasive, non-ablative treatments designed to reverse these biologic processes and restore the disc and facet functioning may become a reality.

### **Intervertebral Disc**

Intervertebral disc degeneration is a major cause of musculoskeletal disability in humans<sup>8-10</sup>. Degeneration has been linked to low back pain; however, the exact relationship between the two remains uncertain<sup>11,12</sup>. The macroscopic features characterizing disc degeneration include the formation of tears within the annulus fibrosus (AF), progressive fraying and dehydration of the nucleus pulposus (NP) with eventual loss of the anular-nuclear distinction<sup>8,9,13</sup>. These pathologic alterations result in substantial changes in the functioning of the disc. Unquestionably, disc degeneration is a multi-factorial process influenced by genetics, lifestyle conditions (including obesity, occupation, and smoking), biomechanical loading, and biochemical event<sup>14,15</sup>.

#### *Intervertebral Disc Biomechanics*

The disc is capable of converting axial spinal loads into tensile hoop stresses in the outer AF while allowing motion of the vertebral segment. This behavior of the IVD is dependent on the distinct biomechanical properties of the NP and AF. The proteoglycan-

rich NP acts as an internal semi-fluid mass, whereas the collagen-rich AF, acts as a laminar fibrous container<sup>16</sup>. The hydrostatic properties of the disc arise from its high water content which allows it to support such large loads<sup>17,18</sup>.

The NP in a young adult, acts as a viscid fluid under applied pressure, but also exhibits considerable elastic rebound, assuming its original physical state upon release<sup>19</sup>. Whereas a major function of the NP is to resist and redistribute compressive forces within the spine, the major function of the AF is to withstand tension. The unique combination of biochemical and biomechanical properties of the AF and NP, allows the intervertebral disc to absorb and disperse the normal loading forces experienced by the spine<sup>19,20</sup>. When one of these two units, either the AF or NP, is compromised, degenerative changes ensue because of the alteration in mechanical force distribution across the functional spinal unit.

Horst and Brinckmann found that the stress distribution across the intervertebral disc and vertebral end plate depends on the degree of disc degeneration<sup>21</sup>. Under pure compressive and eccentric-compressive loading, the healthy lumbar intervertebral disc demonstrated a uniform stress distribution across the entire end plate area. Severely degenerated discs demonstrated the same uniform shape of stress distribution under compressive loading but a non-uniform stress distribution when loaded eccentrically. The asymmetry of the stress distribution in degenerated discs was found to increase with both angle of inclination and degree of degeneration. The asymmetric stress distribution was presumed to occur because of the relatively solid nature of the degenerated disc and its inability to

conform to the eccentric loads. These results have been further supported by more recent studies as well<sup>22,23</sup>.

With advancing degeneration, it appears that the proportion of load transmission shifts to the posterior elements. Yang and King indirectly measured facet forces by using an intervertebral load cell to measure the load transferred through the disc<sup>11</sup>. The model predicted a significant increase in facet load for segments with degenerated discs. The increase was more prominent as the eccentricity of the applied compressive load increased posteriorly. This biomechanical sequence of disc degeneration leading to posterior element load bearing may in fact be what is observed clinically in that disc degeneration typically precedes facet arthrosis<sup>4</sup>.

Clinically, a common observation is that disc degeneration creates instability of the lumbar spine and, therefore, increases range of motion<sup>24</sup>. The interplay between the intervertebral disc geometric and material properties as well as facet joint competence are important in defining the stability of the involved motion segment<sup>25</sup>. Biomechanical studies suggest that changes in stability with disc degeneration are quite complex. The kinematic behavior of a simulated degenerative model under compressive and shear loading were studied by Frei et al<sup>26</sup>. The authors found greater axial translations under compression in the degenerated model (nucleotomy) compared to the normal disc. In anterior shear, the anterior translation was smaller in the degenerated specimens versus the normal specimens. Anterior shear was accompanied by a significant increase in coupled flexion rotation in the degenerative model, which could explain the

counterintuitive decrease in translation. This was attributed to an increase in facet load in degenerated specimens during anterior shear loading. Fujiwara et al, in addition, found in vitro cadaveric specimens that segmental motion changes were much greater in axial rotation compared to lateral bending, flexion, and extension<sup>27</sup>. Ochia et al also found an increase in torsional and flexion and extension movements in vivo<sup>28</sup>. These kinematic studies ultimately can be related clinically to the concept that excessive motion beyond normal soft tissue or bony constraints causes compression or stretching of the neural elements, or deformation of the soft tissue<sup>29</sup>. These instabilities can cause abnormal motion, contact forces and accelerate facet degeneration and osteoarthritis. Eventually as pointed out by Kirkaldy-Willis, with advancing degeneration the motion segment ultimately becomes less mobile, although the remaining motion may certainly be painful<sup>24</sup>. As the disc becomes less mobile, this may in turn decrease the intrinsic disc strength and may decrease nutrition to the disc<sup>30</sup>.

Besides spinal instability creating degenerative disc disease, another competing biomechanical cause for disc degeneration is the “wear and tear” hypothesis. In this mechanism, a series of minor mechanical trauma to the disc accumulates eventually creating disc weakening. This weakening results in further injury, and a vicious cycle ensues ultimately leading to disc degeneration<sup>31,32</sup>. If this model was the main reason for disc degeneration, a logical assumption would be that heavy physical loading, particularly laborers, would have an elevated risk to disc degeneration. Most studies have shown an association between heavy physical loading and disc degeneration<sup>33-42</sup>; however, a study by Friberg and Hirsch did not find an association between occupational and spine

degeneration radiographically<sup>43</sup>. Other studies as well have not demonstrated a clear association<sup>33,44-48</sup>.

Whatever the biomechanical etiology for disc degeneration, researchers have attempted to define a relationship between biomechanical intervertebral disc alterations and symptomatology. More recently, disc dysfunction associated with axial back pain giving rise to so-called internal disc derangement (IDD) has received considerable attention. Magnetic resonance imaging (MRI) is a valuable diagnostic tool in assessing for IDD<sup>49</sup>. MRI allows determination of the proton density of the disc indicative of the state of hydration and can also identify the presence of annular tears. Aprill and Bogduk described the MRI high intensity zone (HIZ), which they believe to be representative of an annular tear extending to the periphery of the disc<sup>50</sup>. The HIZ can be seen on spin echo T2-weighted images as a high intensity signal located in the substance of the posterior annulus fibrosis. (Fig 2-3) The HIZ, has been suggested as, but by no means confirmed to be, associated with discogenic axial back pain<sup>51,52</sup>.

Modic et al described adjacent bony end-plate changes that occur with degeneration of the intervertebral disc<sup>53,54</sup>. Type 1 changes (decreased signal intensity on T1-weighted spin-echo images and increased signal intensity on T2-weighted images) were identified in 20 patients, type 2 changes (increased signal intensity on T1-weighted images and isointense or slightly increased signal intensity on T2-weighted images) in 77 patients, and type 3 changes (decreased signal on T1 and T2-weighted images) in 16 patients). Histopathologic sections in cases of type 1 change demonstrated disruption and fissuring

of the end plates and vascularized fibrous tissue, type 2 changes demonstrated yellow marrow replacement, and type 3 changes demonstrated loss of marrow and advanced bony sclerosis. These signal intensity changes appear to reflect a spectrum of vertebral body marrow changes associated with degenerative disc disease<sup>53</sup>.

### *Mechanical Treatments*

As disc degeneration progresses, the resulting abnormal motion or instability is believed to be a competent cause of spinal pain, likely related to stretching of soft tissues and stimulation of free nerve endings<sup>24,25,55</sup>. Although a precise understanding of what constitutes spinal “instability” remains elusive, numerous treatments aimed at reducing painful spinal motion have been described. Physical therapy using stabilizing exercises has been proposed as an attempt to re-stabilize the “unstable” spine<sup>56,57</sup>. This approach may be more effective when painful segmental motion is the consequence of injury and dysfunction of the paraspinal muscle system that renders the motion segment biomechanically vulnerable in the neutral zone. The clinical diagnosis is based on the report of pain and the observation of movement dysfunction within the neutral zone and the associated finding of excessive intervertebral motion at the symptomatic level.

Other reported techniques for re-stabilizing the spine include intradiscal therapies such as IDET (Intradiscal Electrothermal Therapy), which purportedly attempt to stiffen the motion segment by altering collagen fibers within the intervertebral disc<sup>58,59</sup>. Histologic studies of IVD material after IDET have reported histologic changes of collagen fibril denaturation in the posterior annulus fibrosis<sup>60</sup>. Another re-stabilization approach involves

the use of posteriorly implanted “dynamic devices” that limit but do not eliminate motion. These devices have been extensively implanted in Europe for select cases of mechanical back pain with “instability”. Total disc replacement which provides axial stability while allowing for motion, is being increasingly used for the treatment of painful disc degeneration.

## **Facet Joint**

### ***Biomechanics***

Facet joints are true synovial articulations and undergo degenerative changes similar to those of OA seen in other synovial joints<sup>11,61</sup>. The facet joints are one of the primary stabilizing structures of the spinal motion segment<sup>62,63</sup>. As the degenerative cascade progresses and anterior column support is lost, the facet joints bears more weight and the fulcrum moves dorsally in order to balance the motion segment<sup>64</sup>. With progressive spinal degeneration, the load-bearing patterns of the facet joints are altered<sup>27</sup>.

Fujiwara et al performed a biomechanical and imaging study of human cadaveric spinal motion segments in order to determine the effect of disc degeneration and facet joint osteoarthritis on the segmental flexibility of the lumbar spine<sup>27</sup>. The authors noted that axial rotation was most affected by disc degeneration. Facet cartilage degeneration, especially thinning of the cartilage, causes capsular ligament laxity, which may allow abnormal motion or hypermobility of the facet joint. The authors noted a significant linear correlation between facet cartilage thinning and disc degeneration in the male

cadavers. Cartilage degeneration appeared to further increase the segmental movements already present in the hypermobile, degenerated disc.

Facetectomy studies have been performed by Sullivan et al in the lumbar spine of immature white rabbits to create a facet-mediated degenerative model<sup>65</sup>. The authors resected the inferior articular process on one side at a selected vertebral level and on the opposite side at the adjacent level. The disc height was decreased at the surgical level in 50% of the discs at 6-months and 74% at 12-months. At 9- to 12-months, the discs showed thinning of the posterior AF, circumferential slits in the peripheral AF and an increased area as well as decreased organization of the NP. The facet joints opposite the facetectomy began to show degeneration at 6-months. The authors concluded that the facet joint protects the intervertebral disc from rotational stresses.

Unquestionably, the facet joint complex has an important role in stabilizing the segmental spinal unit<sup>27,32,62,66,67</sup>. As disc disease progresses, increased stress is applied posteriorly accelerating facet osteoarthritis. The resultant facet joint osteoarthritis is likely to change the segmental spinal motion, altering the mechanical forces experienced by the intervertebral disc.

### **Biological Factors**

Cells residing in the both the AF and NP actively regulate the homeostasis of IVD tissue. These cells maintain a balance between anabolism and catabolism by modulating a variety of substances including cytokines, enzymes, enzyme inhibitors and growth factors

in a paracrine and/or autocrine fashion<sup>13,68-70</sup>. Anabolic regulators include polypeptide growth factors, such as insulin-like growth factor (IGF), transforming growth factor- $\beta$  (TGF- $\beta$ ) and bone morphogenetic proteins (BMPs). Other small molecules such as the synthetic peptide of link proteins have also been reported to be regulators of matrix synthesis<sup>13,70,71</sup>. The catabolic process is also mediated by various enzymes, such as matrix metalloproteinases, aggrecanases, and cytokines<sup>72,73</sup>. The degeneration of an IVD results from an imbalance between the anabolic and catabolic processes, or the loss of steady-state metabolism that is maintained in the normal disc.

This delicate homeostatic balance affects the biomechanics of the IVD as well. A healthy IVD is populated by at least two morphologically distinct cells types<sup>74-79</sup>. The majority of cells are small and round, similar to chondrocytes. The second cell type is thought to be a remnant of the primitive notochord and has a vacuolated appearance and prominent intracellular glycogen deposits. Surrounding these cells is a matrix rich in large aggregating proteoglycans (PGs). This matrix imbibes water allowing the NP to resist compressive forces. With disc degeneration, chondrocytic cells are replaced by fibrocytes synthesizing type I collagen<sup>9</sup>. The baseline synthesis of Type II collagen also declines, altering collagen fiber cross-linking<sup>72,73,80</sup>. Additionally, there is a progressive loss of the PG matrix resulting in IVD dehydration and dessication within the NP. These changes create a weaker biomechanic construct to resist compression and shear forces<sup>81</sup>. Last, an overall decrease in disc cell density with age and degeneration is seen. In studies of human intervertebral discs, Gruber et al reported that apoptosis, or programmed cell death, largely accounts for this depopulation over time, and that interventions which

delay or halt apoptotic cell death may constitute a means of treating degenerative disc disease<sup>74</sup>.

In addition to mediating disc degeneration, biochemical events appear to play a significant role in producing disabling spinal pain<sup>13,81,82</sup>. Biochemical events involved in discogenic pain production appear to include the production and release of inflammatory mediators and cytokines from the disc, vascular ingrowth into annular fissures and the stimulation of free nerve endings in the outermost region of the disc<sup>83-85</sup>.

Studies have suggested nutrition as an important factor in the pathogenesis of disc disease<sup>9</sup>. In order to maintain the steady-state metabolism of cells, the IVD requires proper nutrition, which is accomplished by diffusion of nutrients through the end plates and into the IVD. Trauma, cigarette smoking, and other factors that affect the integrity of the end plates and end-plate vasculature may affect diffusion and disturb the nutrition of the disc cells<sup>86</sup>. Vascular channels in the endplate of the intervertebral disc are particularly vital for maintaining the nutrition of the avascular NP. In degenerative discs, the diffusion capacity decreases creating a lower oxygen tension, decreased pH, and accumulation of catabolic byproducts. Typically, vascular channels at the end-plate proliferate to maintain adequate nutrition of the disc. It has been claimed that the induction of new blood vessels in the in end-plate is facilitated by the activation of enzymes such as matrix metalloproteinases<sup>87</sup>; leading to the belief that with IVD injury the activation of these enzymes is the cause of increasing inflammation within the disc. This inflammation is the harbinger of further degeneration, culminating in a vicious cycle

of accelerated degeneration. There are also reports that these channels ultimately disappear with disc degeneration and eventually become obliterated with calcification<sup>88,89</sup>. Further research using microangiography and immunohistochemical analysis are needed in order to determine if the loss in vascularity at the end plate can be reversed.

Genetic factors play a significant role in the degenerative spinal cascade. A twin study by Sambrook et al examined the hypothesis that disc degeneration has a major genetic component. Spine MRIs were obtained for 86 pairs of monozygotic (MZ) twins and 154 dizygotic (DZ) twins. A substantial genetic influence on disc degeneration was found<sup>90</sup>. Further genetic predispositions to disc degeneration have been suggested by other studies on vitamin-D receptor gene polymorphism<sup>91,92</sup>. The authors noted that in 205 young adults, allelic variation (Tt allele) in the vitamin-D receptor gene was associated with multilevel and severe disc degeneration. Unquestionably, the genetic effect on the disc degeneration cascade requires further analysis.

### *Premise for Biological Therapy*

Current treatment options for degenerative disc disease address its clinical symptom, ie pain, as opposed to the pathophysiological root of the disorder. Furthermore, traditional strategies such as fusion of the involved motion segment are not reliable and may even create instability at adjacent levels or even adjacent level degeneration<sup>93</sup>. In recent years, technologies such as disc replacement, aimed at restoring some degree of motion at the involved segment, while eliminating pain have begun to be studied<sup>94</sup>. However, these motion preserving techniques are appropriate for more advanced stages of spinal

degeneration. With a better understanding of the sequence of biologic and biomechanical events associated with spinal degeneration comes the opportunity for earlier interventions (Fig 4). With early disc and/or facet degeneration, biologic strategies aimed at reversing or retarding the degenerative process are appealing.

Biological therapies can be considered to be structural modifying therapies (those that reverse or retard disc or facet degeneration) and/or symptom modifying therapies (those that provide relief from pain). Various biologic strategies to repair or regenerate the disc have been suggested<sup>5,60,95</sup>. Because the disc has only a limited intrinsic capacity for regeneration, the therapeutic approaches are generally geared towards the enhancement of matrix production by injecting proteins or using gene therapy. Some researchers have begun to increase the intrinsic capacity for regeneration by transplanting cells to the disc to repair the damaged disc matrix<sup>96-98</sup>.

One strategy for preventing, arresting, or reversing intervertebral disc degeneration is to increase the accumulation of the extracellular matrix by enhancing its synthesis and/or inhibiting its degradation through the introduction of biological proteins directly into the IVD. Various candidates exist that fulfill these requirements; however, a complete understanding of all the factors involved is far from being complete. Factors that enhance synthesis include TGF- $\beta$ 1, BMP-2, and BMP-7. In vitro studies have already demonstrated that exogenous application of these growth factors can increase extracellular matrix synthesis by IVD cells<sup>99-102</sup>. In addition to increasing the synthesis of PGs, application of BMP-7 has been shown to increase disc height in normal rabbits and

delay loss of disc height in a lapine model of intervertebral disc degeneration<sup>100,103</sup>.

Blocking the effect of catabolic factors also hold promise. MMP-13, otherwise known as collagenase-3, is recognized to be the most potent degrading enzyme of type II collagen, a principal component of IVD<sup>104,105</sup>. Degradation of the disc collagen in turn alters disc homeostasis and affects the IVD's ability to resist compressive and tensile stresses. For example, activation of MMPs can result in as much as 80% loss of tissue glycosaminoglycan content and destruction of the collagen matrix<sup>106</sup>. These changes result in matrix swelling and decreased mechanical strength of the disc. Tissue inhibitors of matrix metalloproteinases (TIMP) are endogenous inhibitors of MMPs and likely play a crucial role in the regulation of matrix degradation<sup>82,107</sup>. Recent studies have demonstrated that gene transfer of TIMP-1 to NP cells in vitro increased PG synthesis as much as fivefold compared with controls<sup>108</sup>. Despite this promising data, exogenous growth factors have a short half life and affect IVD for a limited amount of time<sup>109</sup>.

To provide a longer sustained response to IVD, the focus of biologics has shifted to gene therapy. In a pathological condition that is chronic in nature, a sustained effect of biological treatments is paramount. Gene therapy directs a target cell to synthesize a desired protein by using a viral or nonviral vector to incorporate a genetic sequence into the host genome<sup>110</sup>. This promising treatment modality has been shown both in vivo<sup>111</sup> and in vitro<sup>112</sup> to up-regulate matrix production by the IVD when cDNA for TGF- $\beta$ 1 was introduced into the disc via an adenoviral vector. Results have also been published demonstrating that the transfer of BMP-2 cDNA to the disc by injection of recombinant adenovirus vector reverses early disc height loss<sup>113</sup>. Type II collagen, the most prominent

collagen in the IVD, synthesis has also been promoted by the transfer of the Sox9 gene<sup>114</sup>. To increase the effect of gene transduction, combination gene therapy with TGF- $\beta$ 1, IGF-1, and BMP-2 revealed an additive effect<sup>115</sup>. These studies hold promise, however, as with other biological treatments, obstacles exist preventing routine use of these techniques in human patients. The safety of using viral vectors for gene transfer first needs to be assessed.

Because intervertebral disc degeneration is associated with the loss of healthy cells, gene therapy may not produce a robust response compared to repopulating the disc with responsive cells. Therefore, another method to reverse IVD degeneration attempted by researcher is the injection of cells. Two potential sources are autologous disc cells and mesenchymal stem cells. The former is less ideal as these cells would have to be harvested intrusively from the patient's own degenerative disc and these cells may be abnormal. Marrow stromal cells, on the other hand, may be an ideal candidate. These cells could be utilized using two different approaches. One is through the injection of pluripotent cells that will differentiate upon injection in vivo to repair nonfunctional tissue or generate new tissue<sup>116</sup>. Another approach is a combination of gene therapy and cell delivery. Pluripotent cells engineered with incorporation of a specific gene reimplanted back into the animal providing healthy cells to repopulate the disc and provide increased production of the desired protein<sup>117,118</sup>.

No matter which biological treatment is utilized, all strategies are dependent on proper nutrition of the cells or tissues in the disc<sup>17</sup>. With advanced degeneration, the supply of

nutrients is disturbed by sclerosis of the endplate. Without ample nutrition, any biological therapy will not work. In these situations, traditional strategies will continue to be the mainstay of treatment. In addition, if the stability of the motion segment is significantly compromised due to severe disc degeneration or facet joint arthropathy, biological treatments will likely fail.

Ultimately, with a better understanding of the sequence of biologic and biomechanical events associated with spinal degeneration, the opportunity for earlier interventions will become evident. With early disc and/or facet degeneration, biologic strategies aimed at reversing or retarding the degenerative process are appealing; a step wise approach to treatment will emerge (Fig 4). In early stages of degeneration, injection of biological factors will likely suffice. As degeneration progresses, the utilization of gene therapy and transplantation of exogenous cells will predominate. Difficulty however arises in deciding which patients with early degeneration will become symptomatic and which may warrant intervention. Perhaps sophisticated genetic profiling or identification of markers of symptomatic degeneration will facilitate these decisions.

## **Conclusion**

Degeneration significantly affects the load-bearing and kinematic behavior of the spine. Additionally, changes at the molecular level are observed as intradiscal proteoglycan and type II collagen content is diminished and MMPs are increased. Over the decades since the degenerative cascade was first presented, we have come to appreciate that spinal

degeneration is the end-result of interplay between subtle alterations in mechanical and biochemical properties of the intervertebral disc and facet joint complex. As we gain further insight into the degenerative cascade, the treatment of symptomatic spinal degeneration may eventually involve a combination of less-ablative reconstructive procedures and biological manipulations.

## Figure Legend

Figure 1: Kirkaldy-Willis schematic demonstrating a proposed mechanism for disc and facet degeneration<sup>1</sup>.

Figure 2: Biology of disc disruption. Ciba Collection (*Frank Netter, CIBA COLLECTION OF MEDICAL ILLUSTRATIONS A Compilation of Paintings on the Normal and Pathologic Anatomy of the Nervous System*)

Figure 3: A sagittal MRI demonstrating a degenerated, collapsed L5-S1 disc space as evidenced by the loss of disc height and decreased T2-signal. The white arrow points to an area along the posterior anulus exhibiting an increased T2 signal representative of a high intensity zone (HIZ).

Figure 4: A schematic depiction of the therapeutic options including biologics and traditional treatments.

## Bibliography

1. Kirkaldy-Willis WH, Wedge JH, Yong-Hing K, J. R. Pathology and pathogenesis of lumbar spondylosis and stenosis. *Spine* 1978;3:319-28.
2. Miller J, Schmatz C, Schultz A. Lumbar disc degeneration: correlation with age, sex, and spine level in 600 autopsy specimens. *Spine* 1988;13:173-8.
3. Paajanen H, Erkintalo M, Parkkola R, Salminen J, Kormano M. Age dependent correlation of low back pain and lumbar disc degeneration. *Arch Orthop Trauma Surg* 1997;116:106-7.
4. Butler D, Trafimow JH, Andersson GB, McNeill TW, Huckman MS. Discs degenerate before facets. *Spine* 1990;15:111-3.
5. An HS, Thonar EJ, Masuda K. Biological repair of intervertebral disc. *Spine* 2003;28:S86-92.

6. Lord SM, Barnsley L, Wallis BJ, Bogduk N. Chronic cervical zygapophysial joint pain after whiplash. A placebo-controlled prevalence study. *Spine* 1996;21:1737-44.
7. Lord SM, Barnsley L, Wallis BJ, McDonald GJ, Bogduk N. Percutaneous radio-frequency neurotomy for chronic cervical zygapophysial-joint pain. *N Engl J Med* 1996;335:1721-6.
8. Battie MC, Videman T, Parent E. Lumbar disc degeneration: epidemiology and genetic influences. *Spine* 2004;29:2679-90.
9. Buckwalter JA. Aging and degeneration of the human intervertebral disc. *Spine* 1995;20:1307-14.
10. Anderson JA. Epidemiological aspects of back pain. *J Soc Occup Med* 1986;36:90-4.
11. Yang K, King A. Mechanism of facet load transmission as a hypothesis for low-back pain. *Spine* 1984;9:557-65.
12. Vanharanta H, Sachs B, Spivey M, Guyer R, Hochschuler S, Rashbaum R, Johnson R, Ohnmeiss D, Mooney V. The relationship of pain provocation to lumbar disc deterioration as seen by CT/discography. *Spine* 1987;12:295-8.
13. Roughley P. Biology of intervertebral disc aging and degeneration: involvement of the extracellular matrix. *Spine* 2004;29:2691-9.
14. Le Maitre C, Freemont A, Hoyland J. Localization of degradative enzymes and their inhibitors in the degenerate human intervertebral disc. *J Pathol* 2004;204:47-54.
15. Lee C, Langrana N. A review of spinal fusion for degenerative disc disease: need for alternative treatment approach of disc arthroplasty? *Spine J* 2004;4:173S-6S.
16. Gruber H, Hanley E. Ultrastructure of the human intervertebral disc during aging and degeneration: comparison of surgical and control specimens. *Spine* 2002;27:798-805.
17. Urban J, McMullin J. Swelling pressure of the lumbar intervertebral discs: influence of age, spinal level, composition, and degeneration. *Spine* 1988;13:179-87.
18. Mulholland R, Sengupta D. Rationale, principles, and experimental evaluation of the concept of soft stabilization. *Eur Spine J* 2002;11:S198-205.
19. Lotz J, Hsieh A, Walsh A, Palmer E, Chin J. Mechanobiology of the intervertebral disc. *Biochem Soc Trans* 2002;30 :853-8.
20. Roughley P, Alini M, Antoniou J. The role of proteoglycans in aging, degeneration and repair of the intervertebral disc. *Biochem Soc Trans* 2002;30:869-74.
21. Horst M, Brinckmann P. 1980 Volvo award in biomechanics. Measurement of the distribution of axial stress on the end-plate of the intervertebral body. *Spine* 1981;6:217-32.
22. McNally D, Adams M. Internal intervertebral disc mechanics as revealed by stress profilometry. *Spine* 1997;17 :66-73.
23. Krang M, Seroussi R, Wilder D, Pope M. Internal displacement distribution from in vitro loading of human thoracic and lumbar spinal motion segments: experimental results and theoretical predictions. *Spine* 1987;12:1001-7.
24. Kirkaldy-Willis W, Farfan H. Instability of the lumbar spine. *Clin Orthop* 1982;165:110-23.
25. Mimura M, Panjabi M, Oxland T, Crisco J, Yamamoto I, Vasavada A. Disc degeneration affects the multidirectional flexibility of the lumbar spine . *Spine* 1994;19:1371-80.
26. Frei H, Oxland T, Nolte L. Thoracolumbar spine mechanics contrasted under compression and shear loading. *J Orthop Res* 2002;20:1333-8.

27. Fujiwara A, Lim T, An H, Tanaka N, Jeon C, Andersson G, Haughton V. The effect of disc degeneration and facet joint osteoarthritis on the segmental flexibility of the lumbar spine. *Spine* 2000;25 :3036-44.
28. Ochia R, Inoue N, Renner S, Lorenz E, Lim T, Andersson G, An H. Three-dimensional in vivo measurement of lumbar segmental motion. *Spine* 2006;31:2073-8.
29. Panjabi M. Low back pain and spinal instability *In: Weinstein, JN Gordon, SL Low Back pain: a scientific and clinical overview* 1996;Rosemont:367-84.
30. Stokes I, Iatridis J. Mechanical conditions that accelerate intervertebral disc degeneration: overload versus immobilization. *Spine* 2004;29:2724-32.
31. Adams M, Bogduk N, Burton K, Dolan P. Biomechanics of back pain *New York* 2002;Churchill Livingstone.
32. Adams M, Freeman B, Morrison H, Nelson I, Dolan P. Mechanical irritation of intervertebral disc degeneration. *Spine* 2000;25:1625-36.
33. Battie M, Videman T, Gibbons L, Fisher L, Manninen H, Gill K. 1995 Volvo Award in clinical sciences. Determinants of lumbar disc degeneration: a study relating life-time exposures and magnetic resonance imaging findings in identical twins. *Spine* 1995;20 :2601-12.
34. Evans W, Jobe W, Seibert C. A cross-sectional prevalence study of lumbar disc degeneration in a working population. *Spine* 1989;14:60-4.
35. Battie M, Videman T, Gibbons L, Manninen H, Gill K, Pope M, Kaprio J. Occupational driving and lumbar disc degeneration: a case-control study. *Lancet* 2002;360:1369-74.
36. Biering-Sorensen F, Hansen F, Schroll M, Runenborg O. The relation of spinal x-ray to low back pain and physical activity among 60-year-old men and women. *Spine* 1985;10:445-51.
37. Kellgren J, Lawrence J. Rheumatism in miners: II: X-ray study. *Br J Industr Med* 1952:197-207.
38. Lawrence J, Molyneux M, Dingwall-Fordyce I. Rheumatism in foundry workers. *Br J Industr Med* 1966;23:42-52.
39. Riibimaki H, Mattsson T, Zitting A, Wickstrom G, Hanninen K, Waris P. Radiographically detectable degenerative changes of the lumbar spine among concrete reinforcement workers and house painters . *Spine* 1990;15:114-9.
40. Sairanen E, Brushaber L, Kaskinen M. Felling work, low back pain and osteoarthritis. *Scand J Work Environ Health* 1981;7:18-30.
41. Videman T, Sarna S, Battie M, Koskinen S, Gill K, Paananen H, Gibbons L. The long-term effects of physical loading and exercise lifestyles on back-related symptoms, disability, and spinal pathology among men. *Spine* 1995;20:699-709.
42. Videman, Simonen R, Usenius J, Osterman K, Battie M. The long-term effects of rally driving on spinal pathology. *Clin Biomech (Bristol, Avon)* 2000;15:83-6.
43. Friberg S, Hirsch C. Anatomical and clinical studies on lumbar disc degeneration. *Acta Orthop Scand* 1949;19:222-42.
44. Savage R, Whitehouse G, Roberts N. The relationship between the magnetic resonance imaging appearance of the lumbar spine and low back pain, age, and occupation in males. *Eur Spine J* 1997;6:106-14.

45. Frymoyer J, Newberg A, Pope M, Wilder D, Clements J, MacPherson B. Spine radiographs in patients with low-back pain: an epidemiological study in men. *J Bone Joint Surg Am* 1984;66:1048-55.
46. Hirsch C. Studies on the pathology of low back pain. *J Bone Joint Surg Br* 1959;41:237-43.
47. Caplan P, Freedman L, Connelly T. Degenerative joint disease of the lumbar spine in coal miners: a clinical and x-ray study. *Arthritis Rheum* 1966;9:693-702.
48. Burns J, Loecker T, Fischer J, Bauer D. Prevalence and significance of spinal disc abnormalities in an asymptomatic acceleration subject panel. *Aviat Space Environ Med* 1996;67:849-53.
49. Narvani A, Tsiridis E, Ishaque M, Wilson L. "Pig Tail" technique in intradiscal electrothermal therapy. *J Spinal Disord Tech* 2003;16:280-4.
50. Aprill C, Bogduk N. High-intensity zone: a diagnostic sign of painful lumbar disc on magnetic resonance imaging. *Br J Radiol* 1992;65:361-9.
51. Lam K, Carlin D, Mulholland R. Lumbar disc high-intensity zone: the value and significance of provocative discography in the determination of the discogenic pain source. *Eur Spine J* 2000;9:36-41.
52. Schellhas K. HIZ lesions. *Spine* 1997;22:1538.
53. Modic M, Steinberg P, Ross J, Masaryk T, Carter J. Degenerative disk disease: assessment of changes in vertebral body marrow with MR imaging *Radiology* 1988;166:193-9.
54. Mitra D, Cassar-Pullicino V, McCall I. Longitudinal study of high intensity zones on MR of lumbar intervertebral discs. *Clin Radiol* 2004;59:1002-8.
55. Niosi C, Oxland T. Degenerative mechanics of the lumbar spine. *Spine J* 2004;4:202S-8S.
56. O'Sullivan P. Lumbar segmental "instability": clinical presentation and specific stabilizing exercise management. *Man Ther* 2000;5:2-12.
57. McGill S. Low back stability: from formal description to issues for performance and rehabilitation. *Exerc Sport Sci Rev* 2001;29:26-31.
58. Lee J, Lutz G, Campbell D, Rodeo S, Wright T. Stability of the lumbar spine after intradiscal electrothermal therapy. *Arch Phys Med Rehabil* 2001;82:120-2.
59. Davis T, Delamarter R, Sra P, Goldstein T. The IDET procedure for chronic discogenic low back pain. *Spine* 2004;29:752-6.
60. Shah R, Lutz G, Lee J, Doty S, Rodeo S. Intradiskal electrothermal therapy: a preliminary histologic study. *Arch Phys Med Rehabil* 2001;82:1230-7.
61. Yang K, An H, Ochia R, Lorenz E, Inoue N. In vivo measurement changes in lumbar facet joint width during torsion. in *51st Annual Meeting of the Orthopaedic Research Society* 2005;Washington, DC.
62. Adams M, Hutton W. Cadaver lumbar intervertebral joints. *Spine* 1980;5.
63. Adams M, Hutton W. The mechanical function of the lumbar apophyseal joints. *Spine* 1983;8:327-30.
64. Panjabi M, Goel V, Oxland T, Takata K, Duranceau J, Krag M, Price M. Human lumbar vertebrae. Quantitative three-dimensional anatomy. *Spine* 1992;17:299-306.
65. Sullivan J, Farfan H, Kahn D. Pathologic changes with intervertebral joint rotational instability in the rabbit. *Can J Surg* 1971;14:71-9.

66. Panjabi M, Oxland T, Yamamoto I, Crisco J. Mechanical behavior of the human lumbar and lumbosacral spine as shown by three-dimensional load-displacement curves. *J Bone Joint Surg Am* 1994;76:413-24.
67. Adams M, Hutton W, Stott J. The resistance to flexion of the lumbar intervertebral joint. *Spine* 1980;5:245-53.
68. Roberts S, Caterson B, Menage J, Evans E, Jaffray D, Eisenstein S. Matrix metalloproteinases and aggrecanase: their role in disorders of the human intervertebral disc. *Spine* 2000;25:3005-13.
69. Oegema T. The role of disc cell heterogeneity in determining disc biochemistry: a speculation. *Biochem Soc Trans* 2002;30:839-44.
70. Oegema T. Biochemistry of the intervertebral disc. *Clin Sports Med* 1993;12:419-39.
71. Cadderton R, Shimer A, Gilbertson L, Kang J. Advances in gene therapy for intervertebral disc degeneration. *Spine J* 2004;4:341S-47S.
72. Mwale F, Demers C, Petit A, Roughley P, Poole A, Steffen T, Aebi M, Antoniou J. A synthetic peptide of link protein stimulates the biosynthesis of collagens II, IX, and proteoglycans by cells of the intervertebral disc. *J Cell Biochem* 2003;88:1202-13.
73. Mwale F, Roughley P, Antoniou J. Distinction between the extracellular matrix of the nucleus pulposus and hyaline cartilage: a requisite for tissue engineering of intervertebral disc. *Eur Cell Mater* 2004;8:58-64.
74. Gruber H, Norton H, Hanley E. Anti-apoptotic effects of IGF-1 and PDGF on human intervertebral disc cells in vitro. *Spine* 2000;25:2153-7.
75. Gruber H, Leslie K, Ingram J, Norton H, Hanley E. Cell-based tissue engineering for the intervertebral disc: in vitro studies of human disc cell gene expression and matrix production within selected cell carriers. *Spine J* 2004;4:44-55.
76. Gruber H, Leslie K, Ingram J, Hoelscher G, Norton H, Hanley E. Colony formation and matrix production by human annulus cells: modulation in three-dimensional culture. *Spine* 2004;29:E267-74.
77. Gruber H, Ingram J, Leslie K, Norton H, Hanley E. Cell shape and gene expression in human intervertebral disc cells: in vitro tissue engineering studies. *Biotech Histochem* 2003;78:109-17.
78. Gruber H, Hanley E. Biologic strategies for the therapy of intervertebral disc degeneration. *Expert Opin Biol Ther* 2003;3:1209-14.
79. Gruber H, Hanley E. Observations on morphologic changes in the aging and degenerating human disc: secondary collagen alterations. *BMC Musculoskelet Disord* 2002;3:9.
80. Pokharna H, Phillips F. Collagen crosslinks in human lumbar intervertebral disc aging. *Spine* 1998;23:1645-8.
81. Fujita K, Nakagawa T, Hirabayashi K, Nagai Y. Neutral proteinases in human intervertebral disc. Role in degeneration and probable origin. *Spine* 1993;18:1766-73.
82. Takahashi M, Hoshino H, Ishihara C, Kushida K, Inoue T. The effect of prostaglandin E1 on human bone metabolism: evaluation by biochemical markers for bone turnover. *Endocr Res* 2000;26:119-28.
83. Kääpä E, Zhang L, Muona P, Holm S, Vanharanta H, Peltonen J. Expression of type I, III, and VI collagen mRNAs in experimentally injured porcine intervertebral disc. *Connect Tissue Res* 1994;30:203-14.

84. Kääpä E, Holm S, Han X, Takala T, Kovanen V, Vanharanta H. Collagens in the injured porcine intervertebral disc. *J Orthop Res* 1994;12:93-102.
85. Kääpä E, Gronblad M, Holm S, Liesi P, Murtomaki S, Vanharanta H. Neural elements in the normal and experimentally injured porcine intervertebral disk. *Eur Spine J* 1994;3:137-42.
86. Cinotti G, Della Rocca C, Romeo S, Vittur F, Toffanin R, Trasimeni G. Degenerative changes of porcine intervertebral disc induced by vertebral endplate injuries. *Spine* 2005;30:174-80.
87. Crean J, Roberts S, Jaffray D, Eisenstein S, Duance V. Matrix metalloproteinases in the human intervertebral discs: role in disc degeneration and scoliosis. *Spine* 1997;22:2877-84.
88. Katz M, Hargens A, Garfin S. Intervertebral disc nutrition. Diffusion versus convection. *Clin Orthop Relat Res* 1986;210:243-5.
89. Holm S, Nachemson A. Nutrition of the intervertebral disc: acute effects of cigarette smoking. An experimental animal study. *Ups J Med Sci* 1988;93:91-9.
90. Sambrook P, MacGregor A, Spector T. Genetic influences on cervical and lumbar disc degeneration: a magnetic resonance imaging study in twins. *Arthritis Rheum* 1999;42:366-72.
91. Videman T, Leppavuori J, Kaprio J, Battie M, Gibbons L, Peltonen L, Koshenvuo M. Intragenic polymorphisms of the vitamin D receptor gene associated with intervertebral disc degeneration *Spine* 1998;23:2477-85.
92. Kawaguchi Y, Kanamori M, Ishihara H, Ohmori K, Matsui H, Kimura T. The association of lumbar disc disease with vitamin-D receptor gene polymorphism . *J Bone Joint Surg Am* 2002;84:2022-8.
93. Ghiselli G, Wang J, Bhatia N, Hsu W, Dawson E. Adjacent segment degeneration in the lumbar spine. *J Bone Joint Surg Am* 2004;86:1497-503.
94. Carl A, Ledet E, Yuan H, Sharan A. New developments in nucleus pulposus replacement technology. *Spine J* 2004;4.
95. Phillips F, Reuben J, FT W. Intervertebral disc degeneration adjacent to a lumbar fusion. An experimental rabbit model. *J Bone Joint Surg Br* 2002;84:289-94.
96. Walsh A, Bradford D, Lotz J. In vivo growth factor treatment of degenerated intervertebral discs. *Spine* 2004;29:156-63.
97. Nishida K, Doita M, Takada T, Shimomura T, Maeno K, Kakutani K, Miyamoto H, Kurosaka M. Biological approach for treatment of degenerative disc diseases. *Clin Calcium* 2005;15:399-406.
98. Moon S, Gilbertson L, Nishida K, Knaub M, Muzzonigro T, Robbins P, Evans C, Kang J. Human intervertebral disc cells are genetically modifiable by adenovirus-mediated gene transfer: implications for the clinical management of intervertebral disc disorders. *Spine* 2000;25:2573-9.
99. Takegami K, An H, Kumano F, Chiba K, Thonar E, Singh K, Masuda K. Osteogenic protein-1 is most effective in stimulating nucleus pulposus and annulus fibrosus cells to repair their matrix after chondroitinase ABC-induced chemonucleolysis *Spine J* 2005;5:231-8.
100. An H, Takegami K, Kamada H, Nguyen C, Thonar E, Singh K, Andersson G, Masuda K. Intradiscal administration of osteogenic protein-1 increases intervertebral disc

height and proteoglycan content in the nucleus pulposus in normal adolescent rabbits. *Spine* 2005;30:25-30.

101. Li J, Kim K, Park J, Elmer W, Hutton W, Yoon S. BMP-2 and CDMP-2: stimulation of chondrocyte production of proteoglycans. *J Orthop Sci* 2003;8:829-35.
102. Thompson J, Oegema T, Bradford D. Stimulation of mature canine intervertebral disc by growth factors. *Spine* 1991;16:253-60.
103. An H, Masuda K. Relevance of in vitro and in vivo models for intervertebral disc degeneration. *J Bone Joint Surg Am* 2006;88:88-94.
104. Reboul P, Pelletier J, Tardif G, Cloutier J, Martel-Pelletier J. The new collagenase, collagenase-3, is expressed and synthesized by human chondrocytes but not by synoviocytes, a role in osteoarthritis. *J Clin Invest* 1996;97:2011-9.
105. Mitchell P, Magna H, Reeves L, Lopresti-Morrow L, Yocum S, Rosner P, Goeghegan K, Hambor J. Cloning expression, and type II collagenolytic activity of matrix metalloproteinase-13 from human osteoarthritic cartilage. *J Clin Invest* 1996;97:761-8.
106. Bonassar L, Stinn J, Paguio C, Frank E, Moore V, Lark M, Sandy J, Hollander A, Poole A, Grodzinsky A. Activation and inhibition of endogenous matrix metalloproteinases in articular cartilage: effects on composition and biophysical properties. *Arch Biochem Biophys* 1996;333:359-67.
107. Handa T, Ishihara H, Ohshima H, Osada R, Tsuji H, Obata K. Effects of hydrostatic pressure on matrix synthesis and matrix metalloproteinase production in the human lumbar intervertebral disc. *Spine* 1997;22:1085-91.
108. Risbud M, Shapiro I, Vaccaro A, Albert T. Stem cell regeneration of the nucleus pulposus. *Spine J* 2004;4:348S-53S.
109. Franceschi R, Wang D, Krebsbach P, Rutherford R. Gene therapy for bone formation: in vitro and in vivo osteogenic activity of an adenovirus expressing BMP7. *J Cell Biochem* 2000;78:476-86.
110. Ratko T, Cummings J, Blebea J, Matuszewski K. Clinical gene therapy for nonmalignant disease. *Am J Med* 2003;115:560-9.
111. Nishida K, Kang J, Gilbertson L, Moon S, Suh J, Vogt M, Robbins P, Evans C. Modulation of the biological activity of the rabbit intervertebral disc by gene therapy: an in vivo study of adenovirus-mediated transfer of the human transforming growth factor beta 1 encoding gene. *Spine* 1999;24:2419-25.
112. Moon S, Nishida K, Gilbertson J, Hall R, Robbins P, Evans C, Kang J. Responsiveness of human intervertebral disc cells to adenovirus mediated transfer of TGF-B1 cDNA in 2D and 3D culture systems: comparison to exogenous TGF-B1. . *Presented at: International Society for the Study of the Lumbar Spine meeting* 2002;Adelaide, Australia.
113. Larson J, Levicoff E, Gilbertson L, Kang J. Biologic modification of animal models of intervertebral disc degeneration. *J Bone Joint Surg Am* 2006;88:83-7.
114. Paul R, Haydon R, Cheng H, Ishikawa A, Nenadovich N, Jiang W, Zhou L, Breyer B, Feng T, Gupta P, He T, Phillips F. Potential use of Sox9 gene therapy for intervertebral degenerative disc disease. *Spine* 2003;28:755-63.
115. Moon S, Nishida K, Gilbertson L, Hall R, Robbins P, Kang J. Biologic response of human intervertebral disc cell to gene therapy cocktail. *Presented at: Orthopedic Research Society meeting* 2001;San Francisco, California.

116. Prockop D. Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* 1997;276:71-4.
117. Shimer A, Chadderdon R, Gilbertson L, Kang J. Gene therapy approaches for intervertebral disc degeneration. *Spine* 2004;29:2770-8.
118. Liu L, Wang G, Lee K, Hoppa N, Buyaner D, Hendricks K, al e. Expression of soluble TNF-RII from transduced human mesenchymal stem cells: in vitro and in vivo efficacy. *Blood* 1999;94.